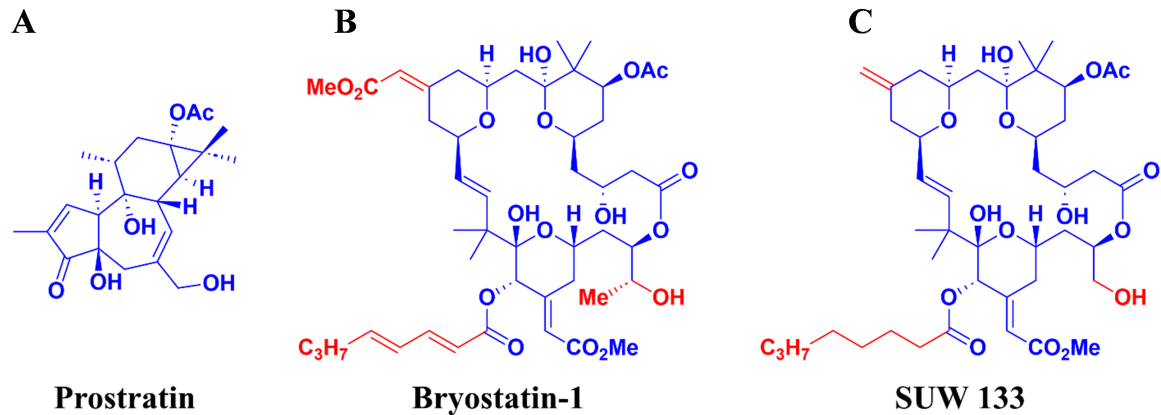
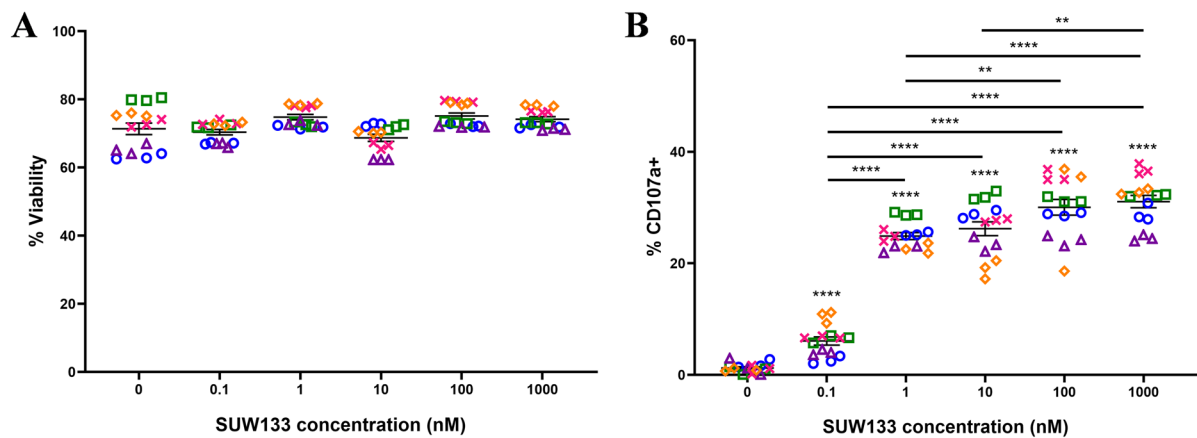


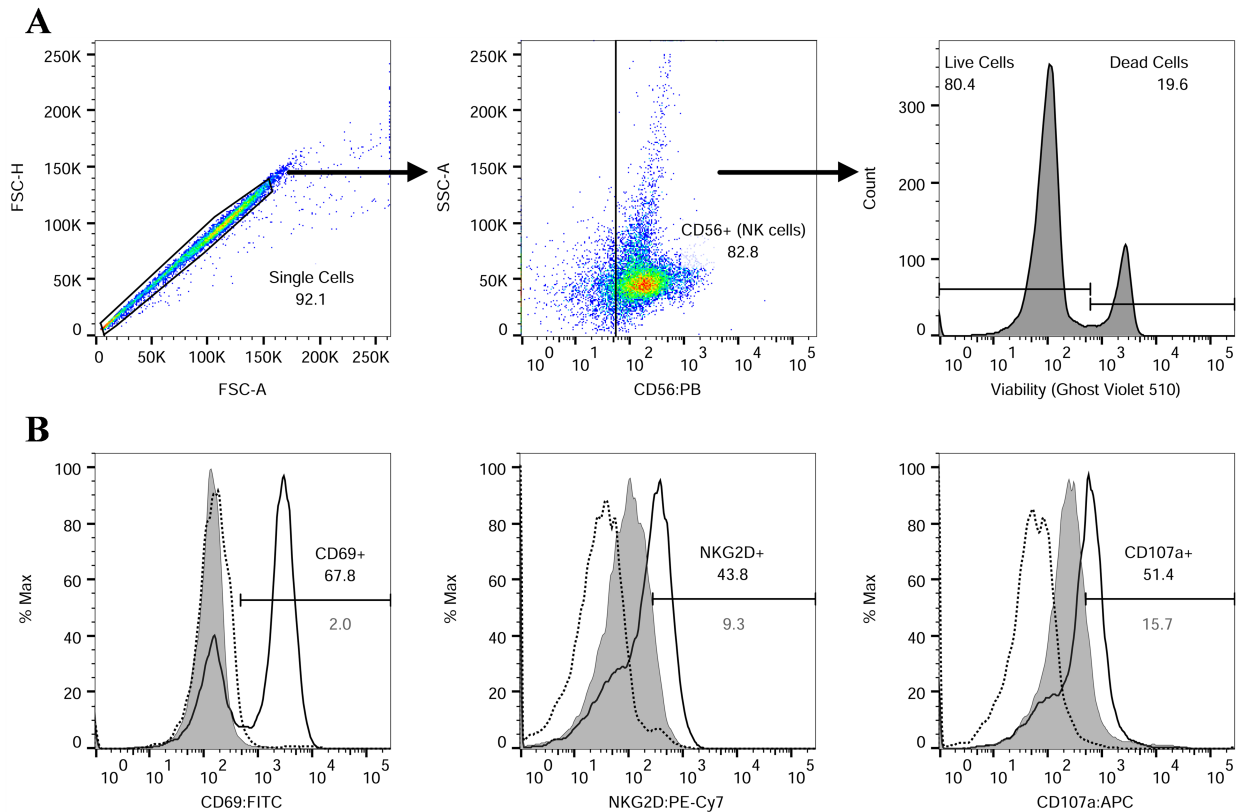
## SUPPLEMENTARY FIGURES



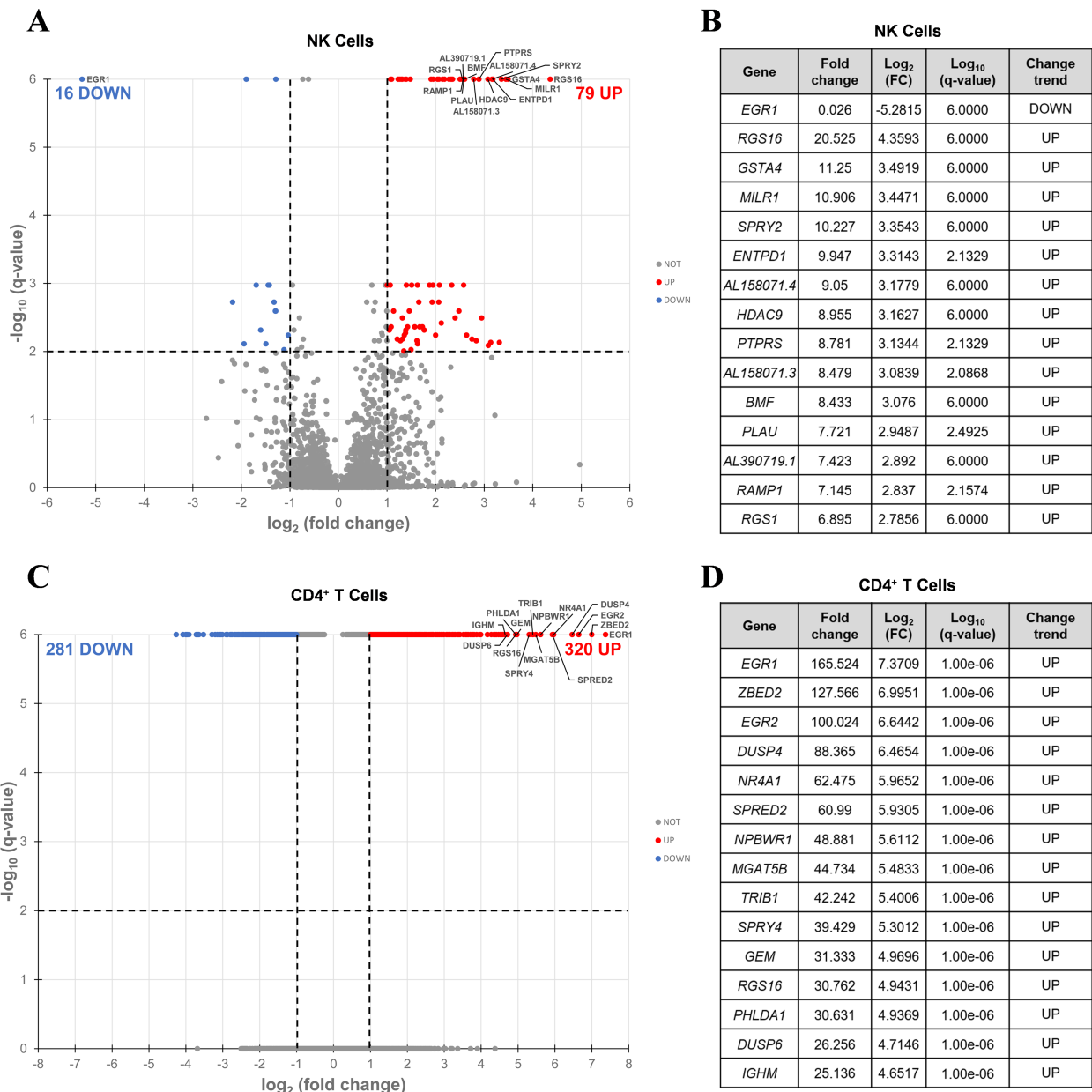
**Supplementary Figure 1. Chemical structure of select PKC modulators.** Chemical structures of prostratin (A), bryostatin-1 (B) and SUW133 (C). Moieties highlighted in red show the differences between bryostatin-1 and SUW133.



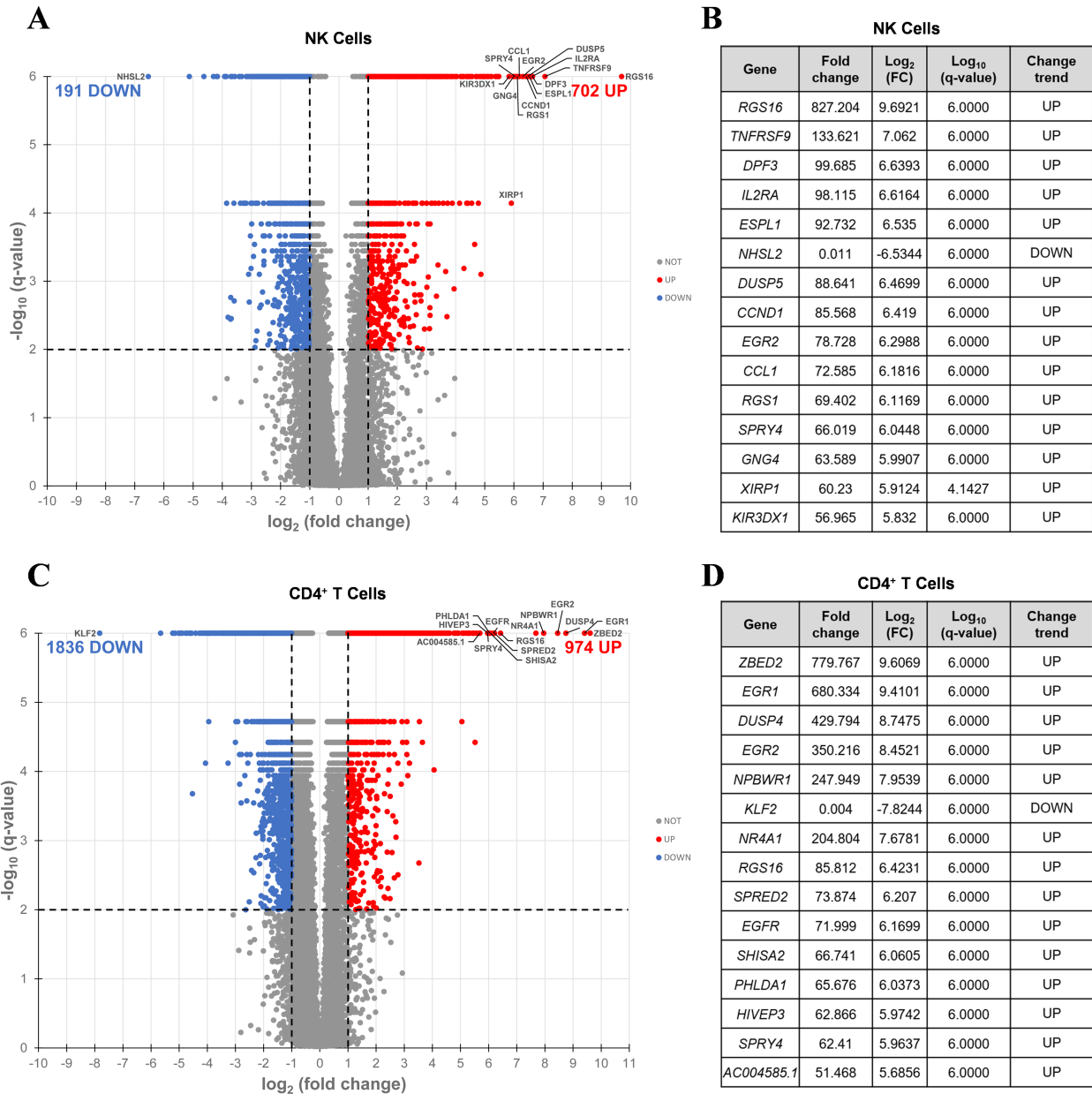
**Supplementary Figure 2. Effect of SUW133 concentration on NK cell viability and degranulation.** NK cells were cultured for 24 hours untreated (DMSO only) or with various concentrations of SUW133 and analyzed for viability (A) and degranulation, as measured by CD107a (B), via flow cytometry. All conditions were conducted in technical triplicates in 5 independent biological replicates (5 donors per condition) resulting in  $n = 15$ . Each bar graph depicts the mean with error bars indicating the standard error of the mean (SEM). A two-tailed, unpaired, unequal variance Student's  $t$ -Test was performed, with (\*\*) indicating  $p < 0.01$  and (\*\*\*\*) indicating  $p < 0.0001$ . The asterisks above each bar graph in panel B correspond to a comparison with the untreated control.



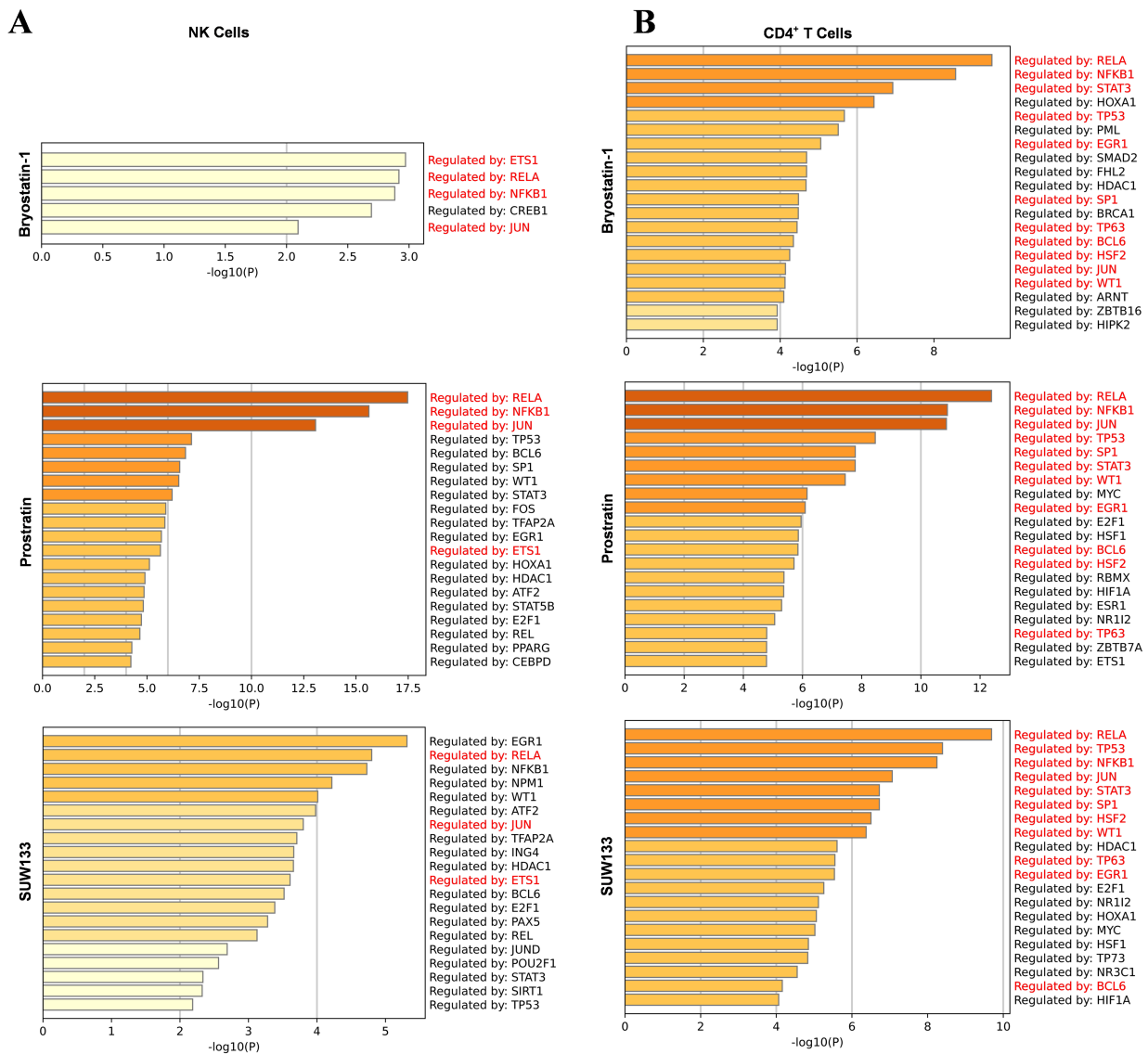
**Supplementary Figure 3. Example flow cytometry profile/gating strategy related to Figure 1.** NK cells were cultured for 24 hours untreated (DMSO only), with 10nM bryostatin-1, 1 $\mu$ M prostratin or 10nM SUW133 and analyzed for viability, CD69, NKG2D and CD107a via flow cytometry. (A) Representative scatter plots and histograms with percentages of gated cells shown. Positive gates were determined using an isotype control. (B) Population comparison of the frequency of CD69<sup>+</sup>, NKG2D<sup>+</sup> CD107a<sup>+</sup> NK cells among untreated (DMSO only; filled gray histograms, gray percentages) and treated cells (black lines, black percentages) along with the isotype control (dotted line) for a representative donor.



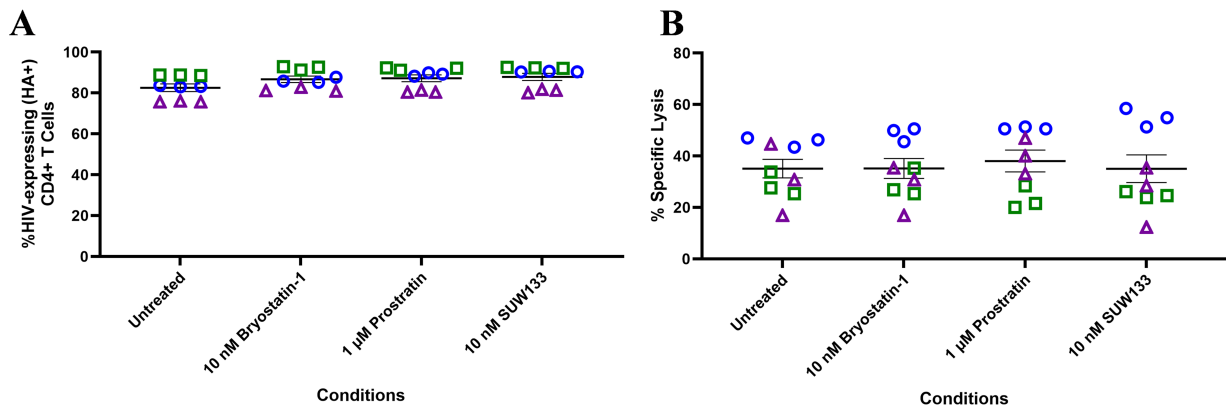
**Supplementary Figure 4. Differentially expressed genes induced by bryostatin-1 in NK and CD4<sup>+</sup> T cells.** Volcano plot of the distribution of all differentially expressed genes in NK cells (A) and CD4<sup>+</sup> T cells (C) treated for 24 hours with 10nM bryostatin-1 compared to the untreated (DMSO only) control. The red and blue dots represent the upregulated and downregulated genes ( $q$ -value < 0.01 and  $|\log_2FC| > 2$ ), respectively. The 15 most differentially expressed genes are labeled on each plot and shown in tables (B) and (D).



**Supplementary Figure 5. Differentially expressed genes induced by prostratin in NK and CD4<sup>+</sup> T cells.** Volcano plot of the distribution of all differentially expressed genes in NK cells (A) and CD4<sup>+</sup> T cells (C) treated for 24 hours with 1 $\mu$ M prostratin compared to the untreated (DMSO only) control. The red and blue dots represent the upregulated and downregulated genes (q-value < 0.01 and  $|\log_2FC| > 2$ ), respectively. The 15 most differentially expressed genes are labeled on each plot and shown in tables (B) and (D).



**Supplementary Figure 6. Summary of enrichment of transcription factors for upregulated genes in NK and CD4<sup>+</sup> T cells treated with PKC modulators.** Metascape gene set enrichment analysis was performed on the lists of upregulated genes for NK cells (A) and CD4<sup>+</sup> T cells (B) treated for 24 hours with 10nM bryostatin-1, 1 $\mu$ M prostratin or 10nM SUW133 using the transcriptional regulatory relationships unraveled by sentence-based text-mining (TRRUST) database. Terms with a  $p < 0.01$ , a minimum count of 3 and an enrichment factor  $>1.5$  were collected and grouped into clusters based on their membership similarities. The  $p$  values were calculated based on the hypergeometric distribution. The data shown is ranked by statistical significance. Terms highlighted in red are shared between all PKC treatment groups.



**Supplementary Figure 7. Impact of PKC modulators on NK cell killing of HIV-infected CD4<sup>+</sup> T cells.** NK cells were pre-treated for 24 hours in vehicle control (DMSO only), with 10nM bryostatin-1, 1μM prostratin or 10nM SUW133 and co-cultured for 4 hours with HIV-infected CD4<sup>+</sup> T cells at a 1:1 effector-to-target (E:T) ratio. All conditions were conducted in technical triplicates in 3 independent biological replicates (3 donors per condition) resulting in n = 9. The percent HIV-expressing target cells (A) and specific lysis (B) was measured, with each color and shape representing results from a different human donor.